NEO-CLERODANE DITERPENOIDS FROM BACCHARIS RHOMBOIDALIS

AURELIO SAN-MARTÍN, JUANA ROVIROSA, CECILIA LABBÉ, ARTURO GIVOVICH, MANUEL MAHÚ* and MARIANO CASTILLO

Departamento de Química and *Departamento de Ciencias Ecológicas, Facultad de Ciencias, Universidad de Chile, Casilla 653, Santiago, Chile

(Received 8 October 1985)

Key Word Index—Baccharis rhomboidalis; Compositae; Astereae; diterpenes; clerodanes; ¹³C NMR.

Abstract—Three new neo-clerodane diterpenoids were isolated together with other known clerodanes from Baccharis rhomboidalis. Their structures were established by spectroscopic methods.

INTRODUCTION

As a continuation of our chemical investigations of the genus *Baccharis* [1], we have studied *B. rhomboidalis* Remy from Central Chile. Besides compounds 1 and 5[2] already isolated from other *Baccharis* species, a new neoclerodane dilactone (7) and three other new clerodane diterpenoids 3, 4 and 6 were found. The new compounds were characterized by spectroscopic methods and some chemical transformations.

RESULTS AND DISCUSSION

Compounds 1-4 were shown to correspond to inseparable mixtures of epimeric cyclic acetals. Compound 3 differed in its ¹H NMR spectrum from that of 1 by the replacement of the tertiary methyl group by a hydroxymethylene group (Table 1). Acetylation of 3 afforded a diacetate, identical in all respects to the natural product 4 also isolated in this work. Comparison of the ¹³C NMR spectra of compounds 1 and 3 (Table 2) indicated C-19 as the site of the additional hydroxyl group in 3 and also confirmed the relative configuration shown in the formulae [1, 3].

Compound 6 was shown to correspond to a clerodanetype diterpene possessing three primary acetoxyl groups, one trisubstituted double bond, two tertiary methyl groups and one secondary methyl group. Again, comparison of the ¹³C NMR spectra of compound 6 with that of 5 (Table 2), also isolated from B. rhomboidalis, showed that the acetyl groups were located at C-15, C-16 and C-18.

The combined UV and IR data of compound 7, $C_{20}H_{26}O_5$, indicated the presence of α,β unsaturated- γ -lactone and hydroxyl groups. Compound 7 was readily acetylated and then oxidized to a ketone which exhibited new IR bands at 1718 and 1680 cm⁻¹, respectively. The ¹H NMR spectrum of 7, showed a one-proton multiplet at $\delta 4.20$ (shifted to $\delta 5.40$ in the acetylated derivative), a narrowly split triplet at $\delta 6.0$ which showed long-range coupling to a 2H doublet, which was characteristic of a proton on the α -carbon of a β -substituted butenolide ring [4]. It also showed signals assigned to an olefinic proton at $\delta 6.50$, a tertiary and a secondary methyl group at $\delta 0.90$ and 0.82, respectively and an AB-system at $\delta 4.62$ and 4.43.

Mild oxidation of 7 afforded an α,β -unsaturated ketone (see Experimental) which showed in its ¹H NMR spectrum a new set of signals at $\delta 6.12$ and 6.50 (dd, 10 and 3.0 Hz) coupled to a triplet at $\delta 3.15$, together with the concomitant disappearance of the methine and olefinic protons at $\delta 4.20$ and 6.50, respectively. These findings were in agreement with the position of the secondary hydroxyl at C-1 in 7 and isomerization of the 3,4-double bond in the newly formed α,β -unsaturated ketone as shown in 9. The axial α -orientation of the 1-hydroxyl group in 7 (and of the 1-acetoxyl group in 8) was suggested by the narrow multiplet ($W_{1/2} = 7.6$) assigned to the equatorial proton at C-1 in these compounds.

The stereochemistry of compound 7 and of its deriva-

Table 1. ¹H NMR data of compounds 3-9 [60 MHz (4 and 7), 100 MHz (8) or 250 MHz (3, 6 and 9), CDCl₃, TMS as int. standard]

Н	3	4	6	7*	8	9†
1				$4.20 \text{ m} \ddagger (W_{1/2} = 7.6)$	$5.40 \text{ m} \ddagger (W_{1/2} = 7.8)$	
3	5.69 t (3.6)	5.73 s (br)	5.53 t (3.0)	6.50 t (4.0) 6.0 m*	6.60 dd (6.0, 4.0) 5.90 m‡	6.50 dd (10.0, 3.1) 5.80 t (1.5)
14				$(W_{1/2} = 4.0)$	$(W_{1/2} = 4.0)$	` ,
15	4.98 t (2.8)* 4.95 d (5.1)	5.07 d (5.0)	4.07 t (6.6)	-,-	,-	
16	3,43 t (7.9) 3.37 t (7.9)	2 07 + (9) 2 25 + (9)	2.06 4 (4.7)	400 4(20)	492 4 (20)	4 60 a (ba)
16′	3.90 t (7.9) 3.40 t (7.9)	3.97 t (8) 3.35 t (8)	3.96 d (4.7)	4.90 d (2.0)	4.82 d (2.0)	4.68 s (br)
17	0.78 d (6.1)	0.79 d (6.0)	0.74 d (5.6)	0.82 d (5.0)	0.89 d (5.0)	0.85 d (5.9)
18			4.42 d (12.8)			
	4.37 s (br)	4.59 s (br)				
18'			4.45 d (12.8)			
19	4.17 d (11.4)	4.48 d (11)		4.62 d (4.0)	4.50 dd (8.0, 1.0)	4.35 d (9.5)
			1.01 s			
19'	3.80 d (11.4)	4.07 d (11)		4.43 d (4.0)	4.38 d (8.0)	4.06 d (br) (9.5)
20	0.72 s	0.74 s	0.67 s	0.90 s	0.78 s	0.90 s
OMc	3.02 s 3.30 s	3.30 s 3.33 s				
Ac		2.02 s	2.01 s (6H)			
			. ,		2.09 s	
		2.04 s	2.02 s (3H)			

^{*}In DMSO-d₆.

Table 2. ¹³C NMR data of compounds 1-9 [20.1 MHz (1, 3, 5, 6) or 62.5 MHz (8, 9), CDCl₃, TMS as int. standard]*

С	1	3	5	6	8	9
1	18.2	17.1	17.1	17.8	65.4	197.2
2	26.5†	26.7†	26.4†	26.5†	34.1	132.6
3	121.1	129.0	125.8	125.8	128.9	137.6
4	147.5	145.0	142.6	142.5	137.1	52.3*
5	37.5	39.1	37.6	37.6	44.6	43.9
6	37.0	30.9	35.9	35.9	32.6	33.8
7	26.2†	26.3†	27.7†	26.9†	27.3	26.5
8	36.1	36.2	35.9	35.9	37.3	37.2
9	38.4	38.5	38.3	38.4	38.9	38.2
10	46.1	46.1	45.8	45.9	48.6	53.0*
11	37.2	37.0	35.3	34.9	35.2	35.6
12	27.2	27.1	29.4	23.7	21.6	23.0
13	37.6	37.3	30.3	31.7	169.6	170.1
14	39.2	42.8	35.2	29.2	114.9	115.5
15	105.5 105.0	105.5 105.0	62.8	62.4	173.3	170.2
16	72.0 71.7	72.3 71.7	19.5	66.0	72.6	72.1
17	15.9	15.7	15.7	15.8	17.7	16.3
18	62.3	64.0	64.8	64.8	168.5	173.3
19	21.2	64.6	70.8	70.8	72.6	73.0
20	18.2	18.7	18.2	18.3	14.8	14.9
OMe	54.8 54.3	54.7 54.3	_	_	_	-
<u>M</u> eCO	_	_	20.9	20.9	20.9	-
MeCO		_	168.9	170.8	1 69 .5	

^{*}Multiplicity were obtained with proton-flip method (APT).

^{†6.12}dd (10.0, 2.8), H-2; 3.15 t (3.0), H-4 and 2.62 s, H-10.

[‡] Values in parentheses are coupling constants in Hz and half-band width of multiplets in Hz.

[†]Interchangeable.

tives was ascertained from the combined evidence of their 1 H and 13 C NMR spectra in comparison with the data of a large group of related clerodanes [5, 6]. Thus, the chemical shifts of the tertiary and secondary methyl groups at C-9 and C-8 respectively were in agreement with the data of compounds having both of these substituents as alpha on a trans-clerodane skeleton [7]. The W coupling exhibited by the α H-19 in 8 and 9 confirmed the α -axial configuration of the C-19 methylene group in agreement with trans clerodanes having an axial methyl group at C-5 and an axial H-6 [8].

The neo-clerodane absolute configuration [9] of compound 7 was established by application of the Horeau method [10] which defined as S the absolute configuration of its C-1 axial alcohol (see Experimental). The absolute configuration of compounds 3 and 6 was not established, but it very probably corresponds to that shown in the formulae since all the clerodane terpenoids isolated from *Baccharis* so far have the neo-clerodane configuration.

EXPERIMENTAL

Plant material. Leaves and top parts of Baccharis rhomboidalis Remy were collected in Cajón del Maipo, Santiago, in December. Voucher specimens are kept at the Herbarium (M.M.), Facultad de Ciencias, Universidad de Chile.

Isolation procedure. Dried and ground plant material (3.2 kg) was percolated at room temp. with petrol (60–80°) (3 × 15 L), then with 95% EtOH (3 × 15 L) for 48 hr. The EtOH extract (400 g) was first partitioned between CHCl₃ and MeOH-H₂O (1:9), then the CHCl₃ solubles were partitioned between petrol and MeOH-H₂O (9:1) to give 110 g aq. MeOH solubles. This extract was fractionated by flash-CC on silica gel eluted with mixtures of increasing polarity of petrol and EtOAc. Compounds 1-8 were isolated after repeated chromatography (silica gel) of suitable fractions and further purified by preparation of the acetylated derivatives.

Compound 3. 120 mg, mp 129–133° (Me₂CO). IR v_{max}^{KBr} cm⁻¹: 3200, 3000–2900, 1490, 1220, 1040; ¹H NMR: Table 1; ¹³C NMR: Table 2; MS m/z (rel. int.): 334 [M – H₂O] ⁺ (14), 321 [M – CH₂O] ⁺ (9), 303 (24), 289 (9), 273 (8.7), 272 (37), 271 (33), 263 (10), 227 (10), 189 (6), 187 (9), 175 (40), 174 (21), 173 (100), 147 (32), 145 (67).

Compound 4. Oil (640 mg). IR $v_{\text{max}}^{\text{dim}}$ cm⁻¹: 1720, 1240; ¹H NMR: Table 1; MS m/z (rel. int.): 376 [M – AcOH]⁺ (4.2), 304 (5.4), 271 (100), 189 (22.2), 187 (41.3), 173 (40.5).

Compound 6. Oil (502 mg). IR $v_{\text{max}}^{\text{film}}$ cm⁻¹: 2950, 1740, 1670, 1240, 1220; ¹H NMR: Table 1; ¹³C NMR: Table 2; MS m/z (rel. int.): 390 [M – AcOH] + (6.3), 375 (1.5), 315 (1.5), 255 (21), 189 (91), 120 (54), 119 (75), 109 (55), 107 (54), 106 (39), 105 (100).

Compound 7. 307 mg, mp 234-235° (MeOH). $[\alpha]_D^{25}$ -60.5 (CHCl₃-MeOH, 4:1; c 0.18). IR v_{max}^{KBr} cm⁻¹: 3485, 3120, 2970, 1755, 1730, 1670, 1645; UV λ_{max} 216 nm (16090); MS m/z (rel. int.): 328.1600 $[C_{20}H_{24}O_4, M-H_2O]$ (7.9), 316.1609

Compound 8. Acetylation of 7 (Ac₂O-pyridine) gave 8, mp 225° (CHCl₃-hexane). IR $\nu_{\rm max}^{\rm EE}$ cm⁻¹: 2950, 1755, 1738, 1718, 1620, 1240; ¹H NMR: Table 1; ¹³C NMR: Table 2.

Compound 9. Compound 8 (100 mg) was oxidized with pyridinium dichromate (2 g) in 6 ml DMF at 0° for 3 hr. Usual workup gave 9 (60 mg) mp 219° (EtOH). UV λ_{max} 230 nm (6130); IR ν_{max}^{KBr} cm⁻¹: 3100, 2970, 1770, 1730, 1685, 1630, 1010, ¹H NMR: Table 1; ¹³C NMR: Table 2; MS m/z (rel. int.): 344.1596 [C₂₀H₂₄O₅] (11), 313 (7), 235 (18), 234 [M - C₆H₆O₂] (100), 233 (28), 189 (14), 161 (12), 112 (22), 111 (83), 107 (28).

Compounds 1, 2 and 5 were identified by comparison of their spectral characteristics with those reported in the lit [2]. The hitherto unreported ¹³C NMR data of compounds 1 and 5 are listed in Table 2.

Application of Horeau method [10] to compound 7. A mixture of (\pm) - α -phenylbutyric anhydride (0.49 mmol) and compound 7 (0.23 mmol) in C_5H_5N (2 ml) was kept at room temp. for 18 hr: usual work-up gave 117 mg α -phenylbutyric acid, $[\alpha]_D = -7.0$ and 96 mg of ester. Optical yield 24%

Acknowledgements—We are indebted to Drs. M. González, Universidad de Rosario, Argentina and D. B. MacLean, McMaster University, Canada, for NMR measurements. This work was supported by D. I. B. (Universidad de Chile), Fondo Nacional de Ciencias (Grant 1060-84) and the Organization of America States.

REFERENCES

- San-Martín, A., Givovich, A. and Castillo, M. (1986) Phytochemistry 25, 264.
- Bohlmann, F., Kramp, W., Jakupovic, J., Robinson, H. and King, R. (1982) Phytochemistry 21, 399.
- Cough, J. L., Guthrie, J. P. and Stothers, J. B. (1972) J. Chem. Soc. Chem. Commun. 979.
- Herz, W., Pilotti, A. M., Soderholm, A. C., Shuhama, I. K. and Vichnewki, W. (1977) J. Org. Chem. 42, 3913.
- Luteizn, J. M., van Veldhuizen, A. and Croot, A. (1982) Org. Magn. Reson. 19, 95.
- Wagner, H., Seitz, R., Lotter, H. and Herz, W. (1978) J. Org. Chem. 43, 3339.
- Sharma, S. C., Tandon, J. S., Porter, B., Raju, M. S. and Wenkert, E. (1984) Phytochemistry 23, 1194.
- Bhacca, N. S. and Williams, D. H. (1964) Applications of NMR Spectroscopy in Organic Chemistry. Illustrations from the Steroid Field, pp. 116–121. Holden-Day, San Francisco.
- Rogers, D., Ünal, G. G., Williams, D. J., Lay, S. V., Sim, G. A., Jochi, B. S. and Rabindranath, K. R. (1979) J. Chem. Soc. Chem. Commun. 97.
- 10. Horeau, A. and Kagan, H. B. (1964) Tetrahedron 20, 2431.